Analysis of biometric, physiological, and biochemical traits to evaluate the cadmium phytoremediation ability of eucalypt plants under hydroponics

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Wastewater reclamation and reuse represent a feasible solution to meet the growing demand for safe water. An environmentally sustainable technology such as phytoremediation is targeted for the reclamation of polluted waters. To this end, the capability of different plant species to tolerate and accumulate pollutants has to be investigated. In this work, eucalypt plants were studied by analysing biometric, physiological, and biochemical parameters related to cadmium (Cd) tolerance and accumulation in two clones (“Velino ex 7” and “Viglio ex 358”) of Eucalyptus camaldulensis Dehn. × E. globulus subsp. bicostata (Maiden, Blakely & J.Simm.) J.B. Kirkp exposed to 50 μM CdSO₄ under hydroponics for three weeks. The results indicated that both eucalypt clones have a valuable tolerance to cadmium, expressed as the tolerance index (Ti). Biometric investigations showed that, regardless of the clone, the metal exposure affected most parameters related to biomass allocation and leaf growth. On the contrary, significant differences were found between the clones with respect to the chlorophyll content and the Chl α to Chl b ratio. These findings were also confirmed from the analysis of chlorophyll fluorescence transient (OJIP) using the JIP test. Cadmium accumulation occurred in both clones and in particular in the roots, with a poor amount of metal reaching the aerial parts, and the Velino clone showed the highest Cd accumulation. The metal uptake ratio and the phytoextraction efficiency highlight a good Cd phytoextraction ability, especially for the Velino clone. The results are discussed taking into account that, in wastewater phytoremediation systems, root biomass can be completely harvested allowing for the removal of the absorbed metal. Finally, the notable tolerance to submergence and the large environmental adaptability of eucalypt suggest that this plant species represents an interesting candidate for the phytoremediation of Cd-polluted wastewaters.

Keywords: Heavy Metals, Metal Tolerance, Wastewater, Forest Plants, Chlorophyll Fluorescence, Rhizofiltration

Introduction

Anthropogenic activities have led to the global degradation of the quality of surface and groundwater, thus representing a severe threat to the environment and human health. In this regard, cadmium (Cd) pollution is of special concern due to the high toxicity of this metal to biota, even at low concentrations, and its high solubility in water. Cadmium has no biological function in plant growth but it is readily taken up and accumulated in the edible parts of crops through the metabolic pathways of essential nutrient elements, especially Zn and Fe, thus entering the human food chain, causing a wide variety of acute and chronic toxic effects (Gallego et al. 2012). Due to the world population increase, the demand for safe water represents a global challenge. Thus, appropriate management of the water resources is required. In this regard, wastewater reclamation and reuse has become an attractive solution to the problem of the water shortage related to crop irrigation. In many countries irrigation with raw wastewater is a commonly utilised agricultural practice, possibly causing the accumulation of metals in the soil and their transfer into food plants (Fatta-Kassinos et al. 2011). A considerable interest in the development of alternative wastewater management strategies, which are less expensive and more environmentally sustainable than conventional technologies, has been increasing. Biological approaches such as phytoremediation, i.e. the use of plants to clean up surface and groundwater contaminated by several pollutants, has received considerable attention because of its cost-effectiveness and environmentally friendly nature (Lu et al. 2011). In this context, interesting perspectives for the uti-
sation of forest plants in the reclamation of polluted waters have been evidenced (Gui
Di Nissim et al. 2014, Pietrini et al. 2015).

Environmental pressures on forests, plant growth, and accumulation by forest plants have been focused on poplars and willows due to their deep and extensive root systems, fast growth, high evapo-transpiration rates, easy management, and low impact on trophic chains (Zachini et al. 2009, Gui
Di Nissim et al. 2014). In contrast, little information is available in the literature about the phytoremediation potential of Eucalyptus spp., in particular for the reclamation of heavy metal contaminated waters.

The Eucalyptus genus includes over 500 species greatly adapted to an extremely wide range of environments and characterised by high genetic variability and economic value. In particular, Eucalyptus species have been reported to cope well with environmental constraints such as severe saline and drought conditions, maintaining high transpiration rates (Harfouche et al. 2014). Moreover, growth responses of eucalypt plants to the presence of heavy metals, both in laboratory and field conditions, have been described (Fine et al. 2013, Mughini et al. 2013), highlighting good perspectives for their utilisation for phytoremediation (Dhillon et al. 2008, Pietrini et al. 2015). To this scope, besides the adaptability to different pedoclimatic conditions, the selection of genotypes characterised by particular traits linked to metal tolerance and accumulation has been suggested (Pievini et al. 2010, Zhivotovsky et al. 2011). In previous studies on forest trees, parameters such as the metal uptake ratio, translocation factor, tolerance index, and Cd content in the plant were highlighted as suitable to select genotypes for Cd tolerance and bioaccumulation (Zachini et al. 2009, Iori et al. 2016). Furthermore, a correlation between physiological, biochemical, and molecular parameters and the phytoremediation capability of forest plants was recently assessed (Gaudet et al. 2011, Iori et al. 2015). In this context, chlorophyll a fluorescence analysis represents a useful tool to analyze the changes at the physiological and biochemical level in the photosynthetic apparatus under variable environmental conditions (Cocozza et al. 2014).

Specifically, OJIP fluorescence transient analysis, known as the JIP test (Strasser et al. 2004), has been developed for the quantification of several phenomenological and biophysical expressions together with the energy flux parameters of photosystem II (PS II), and may be used to assess metal stress in vivo (Singh & Prasad 2015). Up to now, no works dealing with Cd tolerance in eucalypt plants were using the JIP test. This novel approach could be of relevance for a successful screening of plant materials for phytoremediation. Finally, hydroponic screening has been proposed as a rapid and suitable technique for testing the phytoremediation ability of woody plants, especially for metal removal in aqueous matrices (Pietrini et al. 2010, Zhivotovsky et al. 2011).

The aim of this study was to investigate the metal tolerance, accumulation, and organ distribution in two hybrid clones of Eucalyptus camaldulensis Dehnh. × E. globulus subsp. bicostata (Maiden, Blakely & J.Simm.) J.B. Kirkp. exposed to Cd under hydroponics, by analyzing the growth responses, chlorophyll and metal content, and chlorophyll fluorescence parameters in order to evaluate their phytoremoval capabilities for Cd-polluted waters. Due to the remarkable ability to grow under partial submersion and the large environmental adaptability of eucalypt plants, the assessment of the metal phytoremoval ability represents the crucial step in order to exploit this plant material as a bio-tool for phytoremediation. Particularly, the possibility to use this forest species for water reclamation could open up interesting perspectives for the phytodepuration of wastewater for agricultural purposes, especially in countries that are currently experiencing water scarcity.

Materials and methods

Plant material and growth conditions

One-year-old root cuttings of two eucalypt hybrid clones Eucalyptus camaldulensis Dehnn. × E. globulus subsp. bicostata (Mai
den, Blakely & J.Simm.) J.B. Kirkp., named Velino and Viglio (Mughini et al. 2013), were grown during summer under shaded greenhouse in hydroponics with third-strength Hoagland’s nutrient solution, pH 6.5. Homogeneous grown plants were utilised for the experiment. Five plants were assigned to each 9 liter pot filled with 0 (control) or 50 µM CdSO4, in third-strength Hoagland’s nutrient solution, pH 6.5, and cultivated for three weeks in greenhouse under natural photoperiod, with mean temperatures of 22-25 °C (night-day) and relative humidity of 60-70%. The Cd concentration was chosen in accordance with previous studies which evaluated the Cd phytoremediation potential of forest plant species (Zacchini et al. 2009, Pietrini et al. 2010). Pots were provided with a pumping system to avoid oxygen deprivation and the nutrient solution was replaced twice a week to maintain initial metal concentration and prevent nutrient depletion. At the end of the experiment, control and treated plants were harvested and washed with distilled water without damaging the roots. After biometric and physiological measurements, plant organs were separated and dried in an oven at 80 °C until a constant weight.

Biometric parameter evaluation

Biomass data were utilised for the calculation of different biometric parameters. The tolerance index (TI) was calculated as the ratio of the dry weight of plant organs of Cd-treated plants to the dry weight of plant organs of control plants. Leaf area was measured using the Leaf Area Meter Li-3000 (Licor, NE, USA). The specific leaf area (SLA) was calculated as the ratio of the leaf area relative to the leaf mass. The calculation of the organ mass ratio was performed as the ratio of the leaf (LMR), stem (SMR) and root (RMR) biomass to the total plant biomass.

Chlorophyll content determination and chlorophyll fluorescence analysis (JIP test)

Two cm² of the last fully expanded leaf, sampled at the end of the experiment, were extracted in 80% chilled acetone in the dark. After centrifugation at 10,000×g for 10 min, absorbance of the supernatant was measured at 646.8 and 663.2 nm. The content of chlorophyll a and b was estimated as described by Lichtentaler (1987).

At the end of the treatment, the chlorophyll a fluorescence transient (OJIP transients) was measured on the last fully expanded leaves of both eucalypt clones, using a Plant Efficiency Analyser (PEA, Hansatech Instruments Ltd., King’s Lynn, UK). The measurements were performed on leaves that were previously adapted to the dark for 60 minutes for the complete oxidation of the photosynthetic electron transport system, and the fluorescence intensity was measured for 1 s after the application of a saturating light pulse of 3000 µmol m⁻² s⁻¹. The JIP-test was employed to analyse OJIP transient and bioenergetics parameters: size and number of active reaction centre of photosynthetic apparatus (Fv/Fm), efficiency of water splitting complex (Fv/Fo), quantum yield of primary photochemistry (Fv/Fo or Φp), performance index of PS II (PI) were determined as described by Strasser et al. (2004).

Cadmium content analysis

Cadmium concentration was measured using an atomic absorption spectrophotometer (Perkin Elmer, Norwalk, CT, USA) on digested samples. Dried material was milled to a fine powder (Tecator Cemotec 1090 Sample Mill – Tecator, Hoganas, Sweden), weighed and mineralised as reported elsewhere (Iori et al. 2015).

The metal uptake ratio describes the capability of the plant to extract and accumulate Cd and it was calculated as the ratio of the Cd content in the whole plant to the Cd content of the corresponding growth solution (Iori et al. 2013).

The metal translocation index describes the ability of the plant to translocate the metal and it was measured as the ratio of the Cd content of the aerial parts (leaves and stem) to the Cd content of the corresponding roots (Iori et al. 2013).

The metal content in plant parts was calculated by multiplying the dry weight by
the metal concentration.

The metal phytoextraction efficiency (mg g\(^{-1}\)) was calculated according to Wang et al. (2007) as the ratio of the Cd amount in the aerial parts (leaves and stem, mg plant\(^{-1}\)) to the corresponding root biomass (g DW plant\(^{-1}\)).

### Statistical analysis

The data reported refer to a single typical experiment with five replicates. Normally distributed data were processed with a one- or two-way analysis of variance (ANOVA), depending on the number of the factors of variability, using the software SPSS\(^*\) (Chicago, IL, USA). Statistical significance of the differences between means was assessed by Duncan’s test (P \(\leq 0.05\)), unless otherwise stated.

### Results and discussion

Metabolic tolerance is a basic factor affecting the efficiency of the phytoremediation processes, and its evaluation represents a step of the utmost importance in plant screening for phytoremediation purposes (Zacchini et al. 2009). In this study, the ability of both eucalypt clones to grow under Cd exposure was analyzed by calculating the Ti (Tab. 1). On the basis of the total plant dry mass, the Ti revealed a remarkable difference in Cd tolerance between the two clones, resulting in far higher tolerance in the Velino clone. With respect to plant organs, no differences between the two eucalypt clones were detected at the stem and leaf level, whereas at the root level the Velino clone showed a higher tolerance to Cd than the Viglio clone. Overall, a notable tolerance to the metal was observed in the two eucalypt clones. Therefore, according to the scheme proposed by Lux et al. (2004), these clones could be defined as highly tolerant (Velino, Ti > 60) and moderately tolerant (Viglio, Ti between 35 and 60). This finding appears of particular interest given the high Cd concentration of the spiked water used in this work which was set at about 2000 times higher than that allowed by the Italian Law for underground waters.

The variation of the biomass allocation among organs is considered a suitable parameter which contributes to the efficient selection of plants to be used in phytoremediation applications. In this work, control plants showed a similar trend in biomass partitioning (Tab. 2), allocating most of the dry mass in the roots, followed by leaves and stems. Although no differences between the two clones were observed upon metal treatment, Cd exposure caused a decrease in RMR, which was around 65% compared to the control, and a slight increase in SMR and LMR. Consistent with this data, a reduction of the RMR in plants treated with metals was previously reported by Vassilev et al. (1998) for Cd and by Iori et al. (2013) for Ni, as the result of their toxicity effect on plant growth. Root growth is considered a useful parameter for metal tolerance tests, as the root appuratus is the first plant organ in close contact with the metal and it is the primary site of metal toxic action (Lux et al. 2004). The reduction of biomass allocation observed in the roots of both eucalypt clones can be ascribed to the disturbance of micronutrient uptake and oxidative stress induction by the metal, thus leading to the inhibition of root cell division and an increase in the lignification, as suggested by several authors (Vassilev et al. 2005, Zhivotovsky et al. 2011, Stoláriková et al. 2012).

When Cd is taken up by the roots, it may be translocated to the aerial parts of the plant via xylem reaching the leaf appuratus. At the leaf level, the damaging effects caused by Cd are particularly targeted in plants screened for phytoremediation. In fact, an efficient photosynthetic machinery allows plants to maintain an effective transpiration flux that drives metal translocation to the aerial parts (Pietrini et al. 2010). In the present study, the effect of Cd on the leaves of eucalypt plants was analysed by measuring biometric and biochemical parameters such as the total leaf area, the SLW and SLA (Tab. 3), and the chlorophyll content (Tab. 4). Concerning the total leaf area, differences between the two clones were observed in the control plants, and this parameter was higher in the Viglio clone. Consistent with the findings of several authors (Vassilev et al. 1998, Zacchini et al. 2009), even though on different plant species, the Cd treatment caused a reduction of the leaf area in the eucalypt clones, but no differences between the two clones were observed. Contrarily, Cocozza et al. (2015) reported a lack of modification of the leaf area in a poplar clone exposed to Cd. Regarding the SLW, irrespective of the clone, the Cd exposure reduced the mass of the leaf unit, as previously reported by Sharma et al. (2008) in carrot plants exposed for a long time period to Cd and Zn. Regarding the SLA, no differences between the two clones were observed in the control plants, however under Cd treatment this parameter differed significantly between the two clones, resulting in a higher SLA for the Viglio clone. No differences between the control and treated plants were observed in the Velino clone, confirming the reduced sensitivity to Cd.

| Tab. 1 | Tolerance index (Ti) in plant organs and total plant of two eucalypt hybrid clones treated in hydroponics for three weeks with 50 µM CdSO\(_4\). Data are mean values ± standard error (n=5). Different letters in the same column indicate significantly different values after t-test (P ≤ 0.05).
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<tbody>
<tr>
<td>Clone</td>
<td>Roots</td>
<td>Stem</td>
<td>Leaves</td>
<td>Total plant</td>
<td></td>
</tr>
<tr>
<td>Velino</td>
<td>0.68 ± 0.08 *</td>
<td>0.72 ± 0.11</td>
<td>0.89 ± 0.14</td>
<td>0.70 ± 0.11 *</td>
<td></td>
</tr>
<tr>
<td>Viglio</td>
<td>0.44 ± 0.14 *</td>
<td>0.59 ± 0.11</td>
<td>0.72 ± 0.16</td>
<td>0.47 ± 0.10 *</td>
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| Tab. 2 | Organ mass ratio (g g\(^{-1}\) DW) in plants of two eucalypt hybrid clones treated with 0 (C) and 50 µM CdSO\(_4\) (Cd) in hydroponics for three weeks. Data are mean values ± standard error (n=5). Different letters in the same column indicate significantly different values after Duncan test. (**): P<0.001; (ns): not significant.
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<tbody>
<tr>
<td>Clone</td>
<td>Treatment</td>
<td>Root:mass</td>
<td>Stem:mass</td>
<td>Leaf:mass</td>
</tr>
<tr>
<td>Velino</td>
<td>C</td>
<td>0.42 ± 0.01</td>
<td>0.27 ± 0.01</td>
<td>0.31 ± 0.01</td>
</tr>
<tr>
<td></td>
<td>Cd</td>
<td>0.29 ± 0.02</td>
<td>0.31 ± 0.01</td>
<td>0.38 ± 0.03</td>
</tr>
<tr>
<td>Viglio</td>
<td>C</td>
<td>0.41 ± 0.02</td>
<td>0.26 ± 0.004</td>
<td>0.32 ± 0.01</td>
</tr>
<tr>
<td></td>
<td>Cd</td>
<td>0.27 ± 0.21</td>
<td>0.34 ± 0.01</td>
<td>0.38 ± 0.41</td>
</tr>
</tbody>
</table>

| Tab. 3 | Leaf biometric parameters in plants of two eucalypt hybrid clones treated with 0 (C) and 50 µM CdSO\(_4\) (Cd) in hydroponics for three weeks. Data are mean values ± standard error (n=5). Different letters in the same column indicate significantly different values after Duncan test. (*) P<0.05; (**) P<0.001; (ns): not significant.
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<tbody>
<tr>
<td>Clone</td>
<td>Treatment</td>
<td>Leaf Area (cm(^2))</td>
<td>Specific Leaf Weight (SLW, g cm(^{-1}))</td>
<td>Specific Leaf Area (SLA, cm(^2) g(^{-1}))</td>
</tr>
<tr>
<td>Velino</td>
<td>C</td>
<td>148.6 ± 17.8</td>
<td>0.0284 ± 0.0026</td>
<td>37.4 ± 1.6 **</td>
</tr>
<tr>
<td></td>
<td>Cd</td>
<td>131.5 ± 9.4</td>
<td>0.0269 ± 0.0031</td>
<td>35.3 ± 1.1 **</td>
</tr>
<tr>
<td>Viglio</td>
<td>C</td>
<td>175.2 ± 17.1</td>
<td>0.0285 ± 0.0042</td>
<td>35.4 ± 1.7 **</td>
</tr>
<tr>
<td></td>
<td>Cd</td>
<td>161.4 ± 16.6</td>
<td>0.0235 ± 0.0052</td>
<td>43.6 ± 3.5 **</td>
</tr>
</tbody>
</table>

This document includes information on the effects of cadmium on eucalypt plants, including changes in growth parameters, biomass allocation, and leaf area. Statistical analyses were performed to determine the significance of these changes, with results indicating differences between control and treated plants. The study highlights the importance of metal tolerance and the efficiency of phytoremediation in reducing the impacts of metal contamination.
The ratio of Fv/Fm is usually used as a stress indicator, representing the maximum quantum yield of PS II photochemistry, while Fv/Fs indicates the size and number of active photosynthetic centers in the chloroplast and, therefore, the photosynthetic strength of the plant. As reported by Mallick & Mohn (2003), a reduction in the Fv/Fm ratio may occur if re-oxidation of the quinone Qa was limited by a decrease or partial block of electron transport from PS II to PS I. Moreover, a decline in the Fv/Fs ratio may indicate a PS II donor side inhibition connected with modification of the thylakoid membrane structure (Krupa & Baszynski 1995) or a reduction of the pool size of QA associated with PS II activity (Krause & Weis 1991). Specifically, as reported by Dan et al. (2000), the reduction of the Fv/Fm ratio in Helianthus annuus plants exposed to Cd indicated a change in the rate of electron transport from PS II to the primary electron acceptors. The Fv/Fs ratio represents the efficiency of the water-splitting apparatus of the PS II and reflects its state (Kriedemann et al. 1985). As reported by Mallick & Mohn (2003), a notable increase in the Fv/Fs suggests that Cd had a severe impact on the water-splitting site, most probably by replacing manganese from the water-splitting apparatus of the oxidizing side, resulting in a disruption of the photosynthetic reactions. Moreover, Gonzalez-Mendoza et al. (2007) suggested that the changes in the chlorophyll fluorescence parameters (Fv/Fm, Fv/Fs, and Fv/Fs) induced by Cd treatment could be the result of a Ca2+ substitution by Cd2+ in the catalytic center of PS II during photoactivation. Therefore, the increase in the Fv/Fs ratio in both clones after Cd treatment suggested that the water-splitting apparatus of PS II was the primary site of action of Cd. To better evaluate the activity of the photosynthetic apparatus under metal treatment, the Performance Index (PIabs) was calculated (Zurek et al. 2014, Singh & Prasad 2015). The PIabs combines three independent functional steps of photosynthesis, such as the density of the reaction centers in the chlorophyll bed, the excitation energy trapping, and the conversion of excitation energy to electron transport into a single multi-parametric expression (Strasser et al. 2004). Consistent with Singh & Prasad (2015), the higher decrease of the PIabs indicated that in the Viglio clone the structure and the function of Fv/Fs were more affected than those of Fv/Fm under Cd stress in the Viglio clone (Kumárová et al. 2010). Since the Chl a/b ratio is an indicator of both the functional pigment equipment and the light adaptation/acclimation of the photosynthetic apparatus, monitoring its decrease can be used as an early warning system for the toxic effects of metal accumulation in plants (Li et al. 2009). In this regard, Cd induced toxicity on eucalypt plants can also be evaluated by investigating the photochemistry of PS II through the analysis of the chlorophyll fluorescence kinetics (JIP test – Tab. 5). Cadmium treatment decreased Fv/Fs (F0/Fm), as well as Fv/Fs and increased Fv/Fm, to the same extent in both clones, highlighting the impairment of PSI1. Comparable results were also found by Singh & Prasad (2015) and by Li et al. (2015) respectively, in Solanum melongena L. and in Esholtzia argyi L. plants exposed to different Cd concentrations, while Cozzoza et al. (2014) observed that the chlorophyll fluorescence was not affected by Cd addition in a poplar clone.

**Tab. 4** Chlorophyll content and chlorophyll ratio evaluated in plants of two eucalypt hybrid clones treated with 0 (C) and 50 µM CdSO4 (Cd) in hydroponics for three weeks. Data are mean values ± standard error (n=5). Different letters in columns indicate significantly different values after Duncan’s test. (*): P ≤ 0.01; (***): P ≤ 0.001; (ns): not significant.

<table>
<thead>
<tr>
<th>Clone</th>
<th>Treatment</th>
<th>Chl a (µg cm⁻²)</th>
<th>Chl b (µg cm⁻²)</th>
<th>Tot Chl (µg cm⁻²)</th>
<th>Chl a / Chl b</th>
</tr>
</thead>
<tbody>
<tr>
<td>Viglio</td>
<td>C</td>
<td>21.08 ± 0.53 a</td>
<td>6.57 ± 0.16 a</td>
<td>27.65 ± 0.60 a</td>
<td>3.2070 ± 0.0005 a</td>
</tr>
<tr>
<td></td>
<td>Cd</td>
<td>14.54 ± 0.60 b</td>
<td>4.55 ± 0.18 b</td>
<td>19.09 ± 0.79 b</td>
<td>3.1890 ± 0.0053 b</td>
</tr>
<tr>
<td>Velino</td>
<td>C</td>
<td>22.79 ± 0.89 a</td>
<td>7.11 ± 0.28 a</td>
<td>29.91 ± 1.17 a</td>
<td>3.2034 ± 0.0016 a</td>
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<tr>
<td></td>
<td>Cd</td>
<td>11.99 ± 0.71 c</td>
<td>3.79 ± 0.21 c</td>
<td>15.79 ± 0.92 c</td>
<td>3.1533 ± 0.0102 c</td>
</tr>
</tbody>
</table>

**Tab. 5** Leaf chlorophyll fluorescence parameters, size and number of active reaction centre of photosynthetic apparatus (Fv/Fm), efficiency of water splitting complex (Fv/Fs), quantum yield of primary photochemistry (Fv/F0), performance index of PS II (PIabs) evaluated in control (C) and in 50 µM CdSO4 (Cd) treated cuttings of two eucalypt hybrid clones in hydroponics for three weeks. Data are mean values ± standard error (n=5). Different letters in columns indicate significantly different values after Duncan’s test. (*): P ≤ 0.05; (***): P ≤ 0.001; (ns): not significant.

<table>
<thead>
<tr>
<th>Clone</th>
<th>Treatment</th>
<th>Fv/Fs</th>
<th>Fv/F0</th>
<th>Fv/F0 (F0/Fm)</th>
<th>PIabs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Viglio</td>
<td>C</td>
<td>3.86 ± 0.08</td>
<td>0.259 ± 0.005</td>
<td>0.793 ± 0.003</td>
<td>13.98 ± 0.72 a</td>
</tr>
<tr>
<td></td>
<td>Cd</td>
<td>2.61 ± 0.09</td>
<td>0.392 ± 0.015</td>
<td>0.719 ± 0.007</td>
<td>4.97 ± 0.39 b</td>
</tr>
<tr>
<td>Velino</td>
<td>C</td>
<td>3.63 ± 0.08</td>
<td>0.277 ± 0.006</td>
<td>0.783 ± 0.003</td>
<td>13.16 ± 0.62 a</td>
</tr>
<tr>
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<td>Cd</td>
<td>2.35 ± 0.10</td>
<td>0.430 ± 0.020</td>
<td>0.701 ± 0.010</td>
<td>2.22 ± 0.21 c</td>
</tr>
</tbody>
</table>

**Tab. 6** Root, stem, leaf and total cadmium content (mg plant⁻¹), uptake ratio (mg g⁻¹), metal translocation index and phytoextraction efficiency (mg g⁻¹) in plants of eucalypt hybrid clones treated in hydroponics for three weeks with 50 µM CdSO4. Data are mean values ± standard error (n=5). Different letters in the same column indicate significantly different values after t-test (P≤0.05).

<table>
<thead>
<tr>
<th>Clone</th>
<th>Cadmium content</th>
<th>Uptake ratio</th>
<th>Metal Translocation Index</th>
<th>Phytoextraction efficiency</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Root</td>
<td>Stem</td>
<td>Leaves</td>
<td>Total plant</td>
</tr>
<tr>
<td>Velino</td>
<td>36.7 ± 3.3 a</td>
<td>0.21 ± 0.01 a</td>
<td>0.011 ± 0.002</td>
<td>38.5 ± 3.43 a</td>
</tr>
<tr>
<td></td>
<td>21.9 ± 2.3 b</td>
<td>0.12 ± 0.01 b</td>
<td>0.006 ± 0.001</td>
<td>23.3 ± 2.51 b</td>
</tr>
</tbody>
</table>
PS II and the activity of the electron-transfer chain are more vulnerable to Cd compared to the Velino clone. 

Metal content is an important parameter to investigate in candidate plants for phytoremediation, as the ability to absorb and accumulate metals in plant tissue is a basic trait for phytorextraction purposes. This trait is also considered a good indicator of the efficient detoxification and tolerance mechanisms (Pietrini et al. 2010). The results regarding the Cd content and distribution among organs of eucalypt clones are shown in Table 6. The two eucalypt clones showed the same pattern of Cd distribution with a preferential metal accumulation in the roots. No differences between the two clones were observed with respect to Cd accumulation in the leaves and stems, whereas at the root level the Velino clone showed a higher Cd accumulation ability than the Viglio clone. Consequently, the total plant Cd content was higher in the clone Viglio compared to the clone Velino. It is worth pointing out that the high metal accumulation in the roots is a very useful trait to exploit in plants selected for water reclamation. In fact, in plant-based water decontamination systems the root biomass can be totally harvested, thus removing the absorbed metal. The accumulation of Cd in the roots of eucalypt plants is in agreement with findings by Gomes et al. (2012) and Fine et al. (2013).

To better highlight the ability of the two clones to extract Cd from the solution and accumulate it in plant tissues, the metal uptake ratio and the phytoreextraction efficiency were evaluated (Tab. 6). In both cases, the data confirmed the greater performance of the Velino clone, since the Cd uptake ratio as well as the phytorextraction efficiency were far higher when compared to the Viglio clone. It is worth underlining that the metal uptake ratio of the Velino clone was slightly higher than that reported by Iori et al. (2015) for willow clones exposed to the same Cd concentration in a similar experiment. With regard to the phytorextraction efficiency, it is noteworthy that, in this experiment, the value of this parameter was 10 times higher than that found by Wang et al. (2007) in plants of Elsholtzia splendens exposed to similar Cd concentrations in soil, and therefore with a different metal bioavailability. Nevertheless, the values for this parameter confirmed the higher bioaccumulation ability of the Velino clone.

Among the physiological processes associated with heavy metal tolerance in plants, the restriction of metal transport to the aerial parts has been highlighted in non hyperaccumulating plants (Lux et al. 2004, Gomes et al. 2012, Iori et al. 2013). In fact, in leaves, Cd represents a very toxic agent, as it interferes with the photosynthetic processes (Pietrini et al. 2005). In this work, the ability of eucalypt plants to translocate the absorbed metal from root to the aerial parts was evaluated by the Cd translocation index (Tab. 6). Consistent with the limited literature present on the matter (Gomes et al. 2012, Fine et al. 2015), the two clones showed a low capability to translocate Cd in the aerial parts, and no differences were observed between the clones with respect to this physiological process.

**Conclusion**

In this work, an investigation into the biochemical, physiological, and biochemical traits associated with the ability of eucalypt plants for the phytoremediation ofCd-polluted waters was performed. The results obtained highlighting the notable ability of this plant species to tolerate and phytorextract the metal from the nutrient solution. Consistent with the little evidence present in the literature, eucalypt plants showed prevalent Cd accumulation in the root system with a low translocation to aerial organs. This finding has remarkable implications for phytoremediation, as the limitation of metal transport to leaves allows plants to better cope with metal exposure and to avoid a recycling of the metal from the rhizosphere to the soil top layers by leaf shedding. For the reclamation of metal polluted waters, the uppermost Cd accumulation in the roots is not to be considered a drawback, as in the water decontamination bio-systems the below-ground plant biomass can be harvested at the end of the decontamination cycle, removing the metal accumulated by the roots. Given the notable tolerance of eucalypt plants to submersion and their adaptability to extremely wide pedoclimatic conditions, the results obtained in this work suggest that this plant species could be proposed for the reclamation of Cd-polluted waters.

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