

# Leaf volatile isoprenoids: an important defensive armament in forest tree species

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Current knowledge on the ecological impact of the emission of volatile isoprenoids by plants is reviewed. This trait is common to many terrestrial species but is scattered across different taxonomic groups; it appears to be related to other ecological traits rather than to phylogenetic relationships. Plants invest high resources to produce volatile isoprenoids, which are likely to play multiple roles in the defence against biotic and abiotic stressors. We describe how constitutive and induced volatile isoprenoids may directly or indirectly defend plants, and briefly address how indirect defence may involve communication with other trophic levels beyond the simple plant-herbivory interaction. It is discussed that, as metabolically costly defensive mechanisms are only activated after attacks, induced volatile isoprenoids may also *prime* other biochemical pathways that are involved in stress resistance responses. It is also surmised that attacked plants may also use volatiles as an airborne communication to *signal* the attack to other leaves or other plant organs, or even other plants, eliciting defence responses.

**Keywords:** Isoprene, Monoterpenes, Plant communication, Plant defence, Stress physiology

## Introduction

Isoprenoids are a group of secondary metabolites produced by plants (Vickers et al. 2009). The isoprenoid common precursor, isopentenyl pyrophosphate (IPP) may be enzymatically transformed into isoprene (the most abundant volatile isoprenoids), or may polymerize with its isoform dimethylallyl pyrophosphate (DMAPP). In the latter case, a range of volatile, semi-volatile, or non-volatile isoprenoids is formed, depending on the molecular weight and on the solubility of these molecules. Volatile isoprenoids (VIPs) are isoprene (C<sub>5</sub>) and monoterpenes (C<sub>10</sub>), which are made by the same chloroplast-based photosynthesis-dependent pathway (the MEP pathway, named from the 2-C-methyl-D-erythritol 4-phosphate intermediate), and homoterpenes (C<sub>11</sub> and C<sub>16</sub>) and

sesquiterpenes (C<sub>15</sub>), which are made in the cytosol from the classic mevalonate (MEV) pathway (Lichtenthaler et al. 1997, Tholl et al. 2011).

This paper will review current understanding on the evolution and ecological roles of volatile isoprenoids, whereas non-volatile isoprenoids, which are known to also have important functions (*e.g.*, the antioxidant function of xanthophylls and carotenoids, see Demming-Adams & Adams 2002), will not be treated here.

VIPs are among the most well known Biogenic Volatile Organic Compounds (BVOCs). Because of their abundance and reactivity, once emitted in the atmosphere volatile isoprenoids play a crucial role in the interaction between biosphere and atmosphere, *e.g.*, entering cycles of ozone and particle formation. This topic has been recently reviewed in great detail by Fowler et al. (2009): we shall focus here on the other multiple ecological roles of volatile isoprenoids, as they have emerged from recent studies. We will mostly examine current knowledge about isoprene and monoterpenes, as forest tree species appear to produce and emit these compounds in large rates compared to other plant species, and their importance in protection against abiotic and biotic stresses is now emerging (Loreto & Schnitzler 2010, Dicke & Baldwin 2010).

Volatile isoprenoids can be emitted by different plant organs. Large and life-long emissions are found in photosynthetic organs (leaves and stems), but periodically large

emissions can be detected from flowers (Knudsen et al. 1993) and roots (Steeghs et al. 2004). Emissions of constitutive blends of monoterpenes, homoterpenes, and sesquiterpenes by flowers are ontogenetically programmed and have an important ecological role in the communication between plants and their hosts, mainly pollinators for cross-fertilization (Heil 2008). These aspects will not be treated here, being the main focus of this paper on foliar emissions of volatile isoprenoids.

Foliar volatile isoprenoids may accumulate in small temporary pools in the leaf mesophyll, or in large pools in specialized organs, *e.g.*, resin ducts (conifers), or oil glands (Lamiaceae). In both cases, volatile isoprenoids may be constitutively emitted or the synthesis and emission may be stress-induced. In the following sections current knowledge on the evolution of constitutive isoprenoids will be reviewed, and ecological considerations about the functions of constitutive and stress-induced isoprenoids will be presented.

## Evolution of constitutive isoprenoids

Emission of isoprenoids takes place in most groups of land plants: dicots, monocots, gymnosperms, pteridophytes and mosses (Kesselmeier & Staudt 1999). Nonetheless, distribution of emitting plants is scattered across the phylogeny of land plants, suggesting multiple gains and/or losses of such metabolites. Several hypotheses have been proposed by the scientific community regarding isoprene evolution: the key enzyme isoprene synthase (*IspS*) may either have evolved many times (Harley et al. 1999), or have been lost many times (Hanson et al. 1999). More recently, Lerdau & Gray (2003) suggested that monoterpene synthases forming the same products have evolved independently in widely separated groups (conifers and angiosperms).

Sharkey et al. (2005) explained that, in angiosperms, isoprene synthases form a monophyletic group, but it is not yet possible to assess whether this means evolution from a common ancestral gene or convergence based on function. According to Affek & Yakir (2002), isoprene is a strong antioxidant, and represents a primitive protection mode against singlet oxygen that evolved later into more dedicated radical scavengers. Vickers et al. (2009) also seem to support the idea that isoprene is a primitive trait, and put forward the interesting speculation that isoprene evolved when plants became terrestrial, in order to help cope with high levels of oxygen and strong oxidative stress, which were non-existent in the oxygen-deprived aquatic medium.

Available inventories (*e.g.*, Kesselmeier &

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Staudt 1999) may suggest that the emission of volatile compounds varies between ecologically different plants: isoprene emission characterises mostly hygrophilic trees (poplars, willows, pedunculate oak) and aquatic plants (common reed), whereas monoterpene emitters are mostly ecologically xerophytic. There is no definite evidence yet about such a correlation between the emission trait and the ecological classification of plants. However, one of the first experiments linking taxonomy and functions seems to indicate that the isoprene emission trait is significantly more widespread in hygrophilic tree species of the Italian flora (F. Loreto et al., unpublished).

Indeed, while chemotypes are distinguishable in plant species that store isoprenoids in large pools, e.g., conifers (Hanover 1992, Michelozzi et al. 2008), in large emitters that do not have storage pools there is apparently no relationship between isoprenoids emission and phylogeny. Genera of the same family, or species within the same genus (e.g., *Quercus*), may have different emission patterns, including both isoprene and monoterpene-emitting species, as well as non- or low-emitters (Loreto et al. 1998). In *Quercus suber* (cork oak), - the only species not storing isoprenoids in which the emission of these volatile compounds has been studied at the intraspecific level (Staudt et al. 2004, Loreto et al. 2009) -, different individuals emit different blends of monoterpenes. It may be suggested that these differences are not only caused by adaptation to the environment, but might rather be associated with past genetic isolation of cork oak populations, whose gene pools have been dated to originate several million years before present (Magri et al. 2007). For this reason, Loreto et al. (2009) hypothesised that the emission profiles can be used as markers of intraspecific diversity, thus helping understand patterns of geographic variations associated with adaptation to the environment. Interestingly, however, the high emission of limonene by Portuguese cork oak populations has been proposed to occur because the trait is associated to high cork yield (Loreto et al. 2009). Further studies are needed to unravel the genetic ground of this association between an agriculturally suitable trait and the pattern of isoprenoid formation.

### **Volatile isoprenoids and abiotic stresses**

Almost all abiotic stress factors are able to affect isoprenoid biosynthesis and emission. Since abiotic stresses inhibit photosynthesis without exception, and since volatile isoprenoids are mainly formed by carbon directly derived from photosynthetic carbon metabolism, a concurrent negative effect of abiotic stresses on photosynthesis and isoprenoids emission would be expected. How-

ever, abiotic stresses generally stimulate biosynthesis and emission of constitutive isoprenoids (Fig. 1a). This is due to simultaneous: (i) elicitation of gene expression and increase of transcript levels of the genes involved in volatile isoprenoid biosynthesis; (ii) increasing contribution of carbon coming from the catabolism of photosynthates into the isoprenoid biochemical pathway; (iii) absence of stomatal closure limitation to emission of volatile isoprenoids (Loreto & Schnitzler 2010).

Under environmental constraints that inhibit photosynthesis, the carbon released as volatile isoprenoids often becomes larger than the photosynthetic carbon uptake, and the foliar carbon budget becomes negative (Brilli et al. 2007). When this occurs at the forest ecosystem level, the amount of carbon fixed by vegetation is reduced, thus strongly influencing both net ecosystem (NEP) and biome (NBP) productivity (Kesselmeier et al. 2002).

The search for the ecological reason and physiological mechanism(s) behind the observed elicitation of isoprenoid production (especially isoprene) under abiotic stress conditions (especially high temperatures and high oxidative conditions) has led to significant research in the past two decades (reviewed by Loreto & Schnitzler 2010).

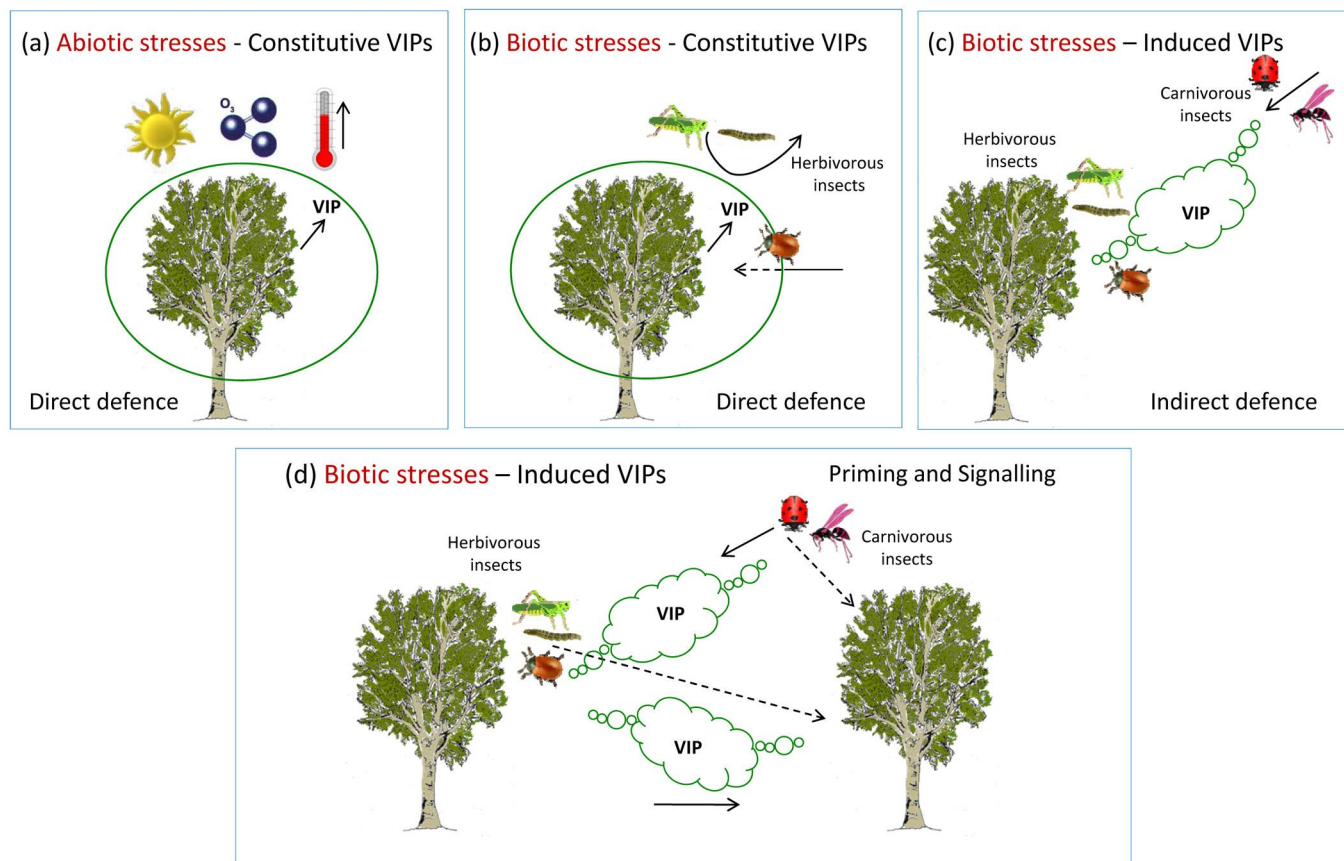
Ecologically, volatile isoprenoids are expected to increase fitness of high-emitting plants, thus accounting for the large metabolic costs of their stress-enhanced biosynthesis. In brief, the two theories that have received prominent experimental support are that: (a) volatile isoprenoids strengthen cellular membranes, thus maintaining the integrity of the thylakoid-embedded photosynthetic apparatus (first postulated by Singsaas et al. 1997); (b) volatile isoprenoids have a more generic antioxidant action, deactivating reactive oxygen species around and inside leaves, thus indirectly reducing oxidation of membrane structures and macromolecules (first proposed by Loreto & Velikova 2001). The two effects of isoprenoids are possibly combined, as explained recently by Loreto & Schnitzler (2010) and Velikova et al. (2012). Yet, experiments with transgenic plants have brought important confirmation that stabilization of membrane integrity may be the primary mechanism that allows maintenance of photosynthetic rates and reduces reactive oxygen species production under stress conditions (Velikova et al. 2011). Behnke et al. (2011) have recently demonstrated that repressing isoprene biosynthesis in high-emitting poplars might provide more carbon for photosynthesis and growth in stress-free environments, although not to the expected extent. However, non-emitting poplars may become more susceptible to herbivores and abiotic stresses. The experiment of Behnke et al. (2011) suggests that isoprene emission

may not be essential for plant survival in temperate habitats, but does not rule out a more significant role of isoprene under stress conditions.

### **Volatile isoprenoids and biotic interactions: direct and indirect defence**

The use of volatile molecules for communication of plants with other organisms is a well-known and fascinating field of study. We have earlier mentioned the use of floral cues to attract pollinators. In this and other insect-plant mutualisms, partners involved play different roles. The sessile partner (the plant) needs to be easily localised and offers rewards (mostly food) to the mobile partner (the insect), who offers a service but also makes its choice in visiting or not a particular individual. This implies that plants have evolved specific traits to be invested in mutualism, whereas insects have not. In fact, the behavioural repertoire utilised by mutualist insects does not differ from that of their relatives not involved in mutualistic associations. Such an asymmetry is particularly obvious in more generalised insect-plant mutualisms (Bronstein et al. 2006).

Plant volatiles are not always useful to the emitting plants (Fig. 1b). Herbivores have evolved the capacity to use volatiles as cues to localize their host. This is typical of constitutive volatiles whose blend is species-specific and is not affected qualitatively by environmental conditions. Numerous examples of such a co-evolution exist in nature, and some of them also apply to forest trees. In poplars, whose adult leaves only emit isoprene at very high rates, the beetle *Chrysomela* (syn.: *Melasoma*) *populi* has learnt that expanding leaves also emit monoterpenes and use this cue to orientate and land on a more rewarding food source (Brilli et al. 2007). Plants also co-evolve, though at a likely slower rate than their animal hosts, and the strategies to defend themselves against herbivores again make use of volatile compounds, including induced isoprenoids. Metabolic changes in plants attacked by herbivores lead to the production of secondary compounds, often named Herbivore-Induced Plant Volatiles (HIPVs), including several volatile isoprenoids that can directly deter herbivores (*direct defence*), or attract herbivores' parasitoids or predators, thus inducing an *indirect induced defence* resulting from the plant interaction with insects of the third trophic level, i.e., carnivores (Fig. 1c - Dicke et al. 2003, Matthes et al. 2010, Dicke & Baldwin 2010). There is again a wealth of examples about direct and indirect defence involving volatile isoprenoids. Volatile isoprenoids contained in the resins of conifers are particularly abundant in young trees where they successfully deter herbivores from feeding (Loreto et al. 2000)



**Fig. 1** - Different roles of volatile isoprenoids (VIPs) in protecting plants from abiotic and biotic stress factors. (a) *Abiotic stresses*. Constitutive VIPs have a protective action against oxidant factors (e.g., ozone), high temperature and other environmental constraints. (b) *Biotic stresses*. Constitutive VIPs repel dangerous herbivores, thus acting as direct defence mechanism. On the other hand, some insects have learnt to use VIPs to locate the host plant, a remarkable example of co-evolution. (c) *Biotic stresses*. The attack by herbivorous insects induces emission of VIPs that attract natural enemies (e.g., carnivorous insects) as an indirect defence mechanism of the host plant. (d) *Biotic stresses and plant communication*. Induced emission of VIPs not only attract natural enemies of herbivorous insects but also help attacked plants communicate to neighbours the presence of risk.

and may even help rapidly sealing mechanical wounds (Pasqua et al. 2002). Although indirect defence has been mostly studied in herbaceous crops, some interactions involving forest trees were also investigated, as reviewed by Holopainen et al. (2009).

Attention has been recently given to even more sophisticated interactions that also involve activation of protection mechanisms through volatile isoprenoid-induced priming or signalling. This is addressed in the next section.

### Volatile isoprenoids and biotic interactions: priming and signalling

Defence mechanisms imply high metabolic costs (Strauss et al. 2002). For this reason plants might have evolved mechanisms to be triggered only as consequence of herbivore attack; this explains why only induced volatile isoprenoids are generally active in the indirect defence. Such strategy may also require *priming* of biochemical mechanisms (mainly activation of stress-induced pro-

teins) preparing plant tissues to react against upcoming attack in a less costly though effective way (e.g., through metabolites with antioxidant and antinutritional properties - Heil & Ton 2010). Priming does not confer resistance *per se*, but makes plants ready for induced resistance when an attack occurs. In this way, it is less costly than a fully implemented defence response (Frost et al. 2008). Induced defence strategies may also require *signalling* - to other leaves or plant organs (e.g., belowground/aboveground), or even other plants - that the infection/infestation has occurred (Baldwin et al. 2006). As for the other defence strategies briefly reviewed above, they may limit the diffusion of herbivore animals, which can quickly move within the plant, or among plants, and cause damages to different organs.

A remarkable example of a coordinated response involving priming and signalling is at the basis of the so called Systemic Induced Resistance (Heil & Ton 2010). Systemic Induced Resistance can be activated in distal organs not yet affected by the attack by two

different mechanisms, both supported by experimental evidence: (i) a systemic transport of (already existing) defensive metabolites; and (ii) a *de novo* expression of resistance mechanism activated by translocated signals from the stressed tissue to distal undamaged tissues. Translocation of signals was traditionally assumed to take place through the vascular system. However, it is now clear that airborne translocation (external transmission of signals by volatiles emitted from the damaged tissues) is also possible and can trigger defensive mechanisms in remote organs both in the same individual and in neighbours (Fig. 1d - Heil & Ton 2010). According to Heil & Ton (2010) there are several reasons explaining why airborne signalling may provide plants with important benefits. First, airborne transportation is a cue that elicits defence responses in those plant parts where resistance is mostly required. Second, signal transmission in the air is faster than the vascular one. Third, airborne and vascular defence signals can act in synergy. Fourth, volatile-mediated signalling

works most efficiently over relatively short distances. In this way, the probability that the leaf nearest to an attacked one belongs to the same plant is fairly high.

Volatile molecules are the best candidates as within- and between-plant signals indicating the presence of herbivores: this info-chemical communication (Frost et al. 2007) implies the aerial transfer of information from a herbivore-attacked plant (the *emitter*) to close growing plants (the *listening receivers*, or *eavesdropping* plants). By eavesdropping HIPVs released from infested neighbors, phenotypes may be tailored to increase fitness. Selection does not favour plants passing important information to competitors (Baldwin et al. 2006). Therefore, eavesdroppers may take advantage of within-plant signalling mechanisms of infected/infested neighbouring HIPVs emitters (Heil & Ton 2010), thus increasing their own fitness.

In fact, within-plant communication may be more effective than plant-to-plant communication in nature. Distance is a crucial parameter in plant-to-plant signalling, and positive reactions are restricted to close surroundings of the emitters. The distance over which airborne cues can affect other plants strongly depends on wind speed, air humidity, and temperature, but in general it appears to be very short (Heil & Ton 2010), especially when emitted volatiles are very reactive, as is the case for most volatile isoprenoids. These compounds also rapidly react with atmospheric pollutants, especially ozone and NO<sub>x</sub>, rapidly disappearing in polluted environments (McFrederick et al. 2008, Blande et al. 2010).

The high reactivity of volatile isoprenoids also makes it possible other, more indirect, priming mechanisms. By rapidly reacting with NO (Velikova et al. 2008) and H<sub>2</sub>O<sub>2</sub> (Loreto & Velikova 2001), isoprene (and perhaps also other volatile isoprenoids) may indirectly quench the two above compounds that are responsible for signalling hypersensitive responses at cellular level (Wilson et al. 2008). The low hypersensitive response may explain the low damage observed in ozonated leaves in an isoprene-rich environment (Loreto et al. 2001). Finally, preliminary results indicate that heavy emitters of volatile isoprenoids may have higher - yet not pathological - levels of these signalling molecules (especially H<sub>2</sub>O<sub>2</sub>) when grown under non-stressed conditions. This fine metabolic adjustment may indicate an additional form of priming able to immunize plants against forthcoming stresses. Further studies are needed to throw light on the above mechanisms, whose impacts on the physiology of stress resistance may be very relevant.

In summary, while recent developments indicate that volatile isoprenoids may not be essential for plant survival in unstressed con-

ditions, an outstanding body of information is now available confirming the multiple roles of these molecules in plant protection against stresses. As environmental and man-made changes (including air pollution and land-use changes) largely affect the capacity to produce and emit biogenic volatile isoprenoids, they may also disturb the defensive and communication performances of plants. The opportunity of producing genetically transformed plants that do not emit volatile compounds ("mute" emitters) and that do not receive these volatile signals ("deaf" plants) represents an extraordinary tool for assessing the importance in nature of volatile isoprenoid signalling and whether they may enhance the plant fitness both in natural communities (Baldwin et al. 2006) and in response to current and future climate change.

## References

- Affek HP, Yakir D (2002). Protection by isoprene against singlet oxygen in leaves. *Plant Physiology* 129: 269-277. - doi: [10.1104/pp.010909](https://doi.org/10.1104/pp.010909)
- Baldwin IT, Halitschke R, Paschold A, von Dahl CC, Preston CA (2006). Volatile signaling in plant-plant interactions: "Talking Trees" in the genomics era. *Science* 311: 812-815. - doi: [10.1126/science.1118446](https://doi.org/10.1126/science.1118446)
- Behnke K, Grote R, Bruggemann N, Zimmer I, Zhou G, Eloheid M, Janz D, Polle A, Schnitzler J-P (2011). Isoprene emission-free poplars - a chance to reduce the impact from poplar plantations on the atmosphere. *New Phytologist* (early view). - doi: [10.1111/j.1469-8137.2011.03979.x](https://doi.org/10.1111/j.1469-8137.2011.03979.x)
- Blande JD, Holopainen JK, Li T (2010). Air pollution impedes plant-to-plant communication by volatiles. *Ecology Letters* 13: 1172-1181. - doi: [10.1111/j.1461-0248.2010.01510.x](https://doi.org/10.1111/j.1461-0248.2010.01510.x)
- Brilli F, Barta C, Fortunati A, Lerdau M, Loreto F, Centritto M (2007). Response of isoprene emission and carbon metabolism to drought in white poplar (*Populus alba*) saplings. *New Phytologist* 175: 244-254. - doi: [10.1111/j.1469-8137.2007.02094.x](https://doi.org/10.1111/j.1469-8137.2007.02094.x)
- Bronstein JL, Alarcón R, Geber M (2006). The evolution of plant-insect mutualisms. *New Phytologist* 172: 412-428. - doi: [10.1111/j.1469-8137.2006.01864.x](https://doi.org/10.1111/j.1469-8137.2006.01864.x)
- Demming-Adams B, Adams WW (2002). Antioxidants in photosynthesis and human nutrition. *Science* 298: 2149-2153. - doi: [10.1126/science.1078002](https://doi.org/10.1126/science.1078002)
- Dicke M, Baldwin IT (2010). The evolutionary context for herbivore-induced plant volatiles: beyond the "cry for help". *Trends in Plant Science* 15: 167-175. - doi: [10.1016/j.tplants.2009.12.002](https://doi.org/10.1016/j.tplants.2009.12.002)
- Dicke M, de Boer JG, Höfte M, Rocha-Granados MC (2003). Mixed blends of herbivore-induced plant volatiles and foraging success of carnivorous arthropods. *Oikos* 101: 38-48. - doi: [10.1034/j.1600-0706.2003.12571.x](https://doi.org/10.1034/j.1600-0706.2003.12571.x)
- Fowler D, Pilegaard K, Sutton MA, Ambus P, Raivonen M, Duyzer J, Simpson D, Fagerli H, Fuzzi S, Schjoerring JK, Granier C, Nefel A, Isaksen ISA, Laj P, Maione M, Monks PS, Burkhardt J, Daemmgen U, Neirynek J, Perronne E, Wichink-Kruit R, Butterbach-Bahl K, Flechard C, Tuovinen JP, Coyle M, Gerosa G, Loubet B, Altimir N, Gruenhage L, Ammann C, Cieslik S, Paoletti E, Mikkelsen TN, Ro-Poulsen H, Cellier P, Cape JN, Horvath L, Loreto F, Ninemets U, Palmer PI, Rinne J, Misztal P, Nemitz E, Nilsson D, Pryor S, Gallagher MW, Vesala T, Skiba U, Bruggemann N, Zechmeister-Boltenstern S, Williams J, O'Dowd C, Facchini MC, de Leeuw G, Flossman A, Chaumerliac N, Erisman JW (2009). Atmospheric composition change: ecosystems-atmosphere interactions. *Atmospheric Environment* 43: 5193-5267. - doi: [10.1016/j.atmosenv.2009.07.068](https://doi.org/10.1016/j.atmosenv.2009.07.068)
- Frost CJ, Appel HM, Carlson JE, De Moraes CM, Mescher MC, Schultz JC (2007). Within-plant signalling via volatiles overcomes vascular constraints on systemic signalling and primes responses against herbivores. *Ecology Letters* 10: 490-498. - doi: [10.1111/ele.2007.10.issue-6](https://doi.org/10.1111/ele.2007.10.issue-6)
- Frost CJ, Mescher MC, Carlson JE, De Moraes CM (2008). Plant defense priming against herbivores: getting ready for a different battle. *Plant physiology* 146: 818-824. - doi: [10.1104/pp.107.113027](https://doi.org/10.1104/pp.107.113027)
- Hanover JW (1992). Applications of terpene analysis in forest genetics. *New Forests* 6: 159-178. - doi: [10.1007/BF00120643](https://doi.org/10.1007/BF00120643)
- Hanson DT, Swanson S, Graham L E, Sharkey TD (1999). Evolutionary significance of isoprene emission from mosses. *American Journal of Botany* 86: 634-639. - doi: [10.2307/2656571](https://doi.org/10.2307/2656571)
- Harley PC, Monson RK, Lerdau MT (1999). Ecological and evolutionary aspects of isoprene emission from plants. *Oecologia* 118: 109-123. - doi: [10.1007/s0044200050709](https://doi.org/10.1007/s0044200050709)
- Heil M, Ton J (2010). Systemic resistance induction by vascular and airborne signalling. *Progress in Botany* 71: 279-306. - doi: [10.1007/978-3-642-02167-1](https://doi.org/10.1007/978-3-642-02167-1)
- Heil M (2008). Indirect defence via tritrophic interactions. *New Phytologist* 178: 41-61. - doi: [10.1111/j.1469-8137.2007.02330.x](https://doi.org/10.1111/j.1469-8137.2007.02330.x)
- Holopainen JK, Heijari J, Nerg AM, Vuorinen M, Kainulainen P (2009). Potential for the use of exogenous chemical elicitors in disease and insect pest management of conifer seedling production. *The Open Forest Science Journal* 2: 17-24. - doi: [10.2174/1874398600902010017](https://doi.org/10.2174/1874398600902010017)
- Kesselmeier J, Staudt M (1999). Biogenic volatile organic compounds (VOC): an overview on emission, physiology and ecology. *Journal of Atmospheric Chemistry* 33: 23-88. - doi: [10.1023/A:1006127516791](https://doi.org/10.1023/A:1006127516791)
- Kesselmeier J, Ciccioli P, Kuhn U, Stefani P, Biesenhal T, Rottenberger S, Wolf A, Vitullo M, Valentini R, Nobre A, Kabat P, Andreae MO (2002). Volatile organic compound emissions in relation to plant carbon fixation and the terrestrial carbon budget. *Global Biogeochemical Cycles* 6: 1126. - doi: [10.1029/2001GB001813](https://doi.org/10.1029/2001GB001813)
- Knudsen JT, Tollsten L, Bergstorm LG (1993). Floral scents: a checklist of volatile compounds

- isolated by head-space techniques. *Phytochemistry* 33: 253-280. - doi: [10.1016/0031-9422\(93\)85502-1](https://doi.org/10.1016/0031-9422(93)85502-1)
- Lerdau M, Gray D (2003). Ecology and evolution of light-dependent and light-independent phyto-genic volatile organic carbon. *New Phytologist* 157: 199-211. - doi: [10.1046/j.1469-8137.2003.00673.x](https://doi.org/10.1046/j.1469-8137.2003.00673.x)
- Lichtenthaler HK, Schwender J, Disch A, Rohmer M (1997). Biosynthesis of isoprenoids in higher plant chloroplasts proceeds via a mevalonate-independent pathway. *FEBS Letters* 400: 271-274. - doi: [10.1016/S0014-5793\(96\)01404-4](https://doi.org/10.1016/S0014-5793(96)01404-4)
- Loreto F, Schnitzler JP (2010). Abiotic stresses and induced BVOCs. *Trends in Plant Science* 15: 154-166. - doi: [10.1016/j.tplants.2009.12.006](https://doi.org/10.1016/j.tplants.2009.12.006)
- Loreto F, Velikova V (2001). Isoprene produced by leaves protects the photosynthetic apparatus against ozone damage, quenches ozone products, and reduces lipid peroxidation of cellular membranes. *Plant Physiology* 127: 1781-1787. - doi: [10.1104/pp.010497](https://doi.org/10.1104/pp.010497)
- Loreto F, Ciccioli P, Brancaleoni E, Valentini R, De Lillis M, Csiky O, Seufert G (1998). A hypothesis on the evolution of isoprenoid emission by oaks based on the correlation between emission type and *Quercus* taxonomy. *Oecologia* 115: 302-305. - doi: [10.1007/s004420050520](https://doi.org/10.1007/s004420050520)
- Loreto F, Nascetti P, Graverini A, Mannozi M (2000). Emission and content of monoterpenes in intact and wounded needles of the Mediterranean Pine, *Pinus pinea*. *Functional Ecology* 14: 589-595. - doi: [10.1046/j.1365-2435.2000.t011-1-00457.x](https://doi.org/10.1046/j.1365-2435.2000.t011-1-00457.x)
- Loreto F, Mannozi M, Maris C, Nascetti P, Ferranti F, Pasqualini S (2001). Ozone quenching properties of isoprene and its antioxidant role in leaves. *Plant Physiology* 126: 993-1000. - doi: [10.1104/pp.126.3.993](https://doi.org/10.1104/pp.126.3.993)
- Loreto F, Bagnoli F, Fineschi S (2009). One species, many terpenes: matching chemical and biological diversity. *Trends in Plant Science* 14: 416-420. - doi: [10.1016/j.tplants.2009.06.003](https://doi.org/10.1016/j.tplants.2009.06.003)
- Magri D, Fineschi S, Bellarosa R, Buonamici A, Sebastiani F, Schirone B, Simeone MC, Vendramin GG (2007). The distribution of *Quercus suber* chloroplast haplotypes matches the palaeogeographical history of the western Mediterranean. *Molecular Ecology* 16: 5259-5266. - doi: [10.1111/j.1365-294X.2007.03587.x](https://doi.org/10.1111/j.1365-294X.2007.03587.x)
- Matthes MC, Bruce TJA, Ton J, Verrier PJ, Pickett JA, Napier JA (2010). The transcriptome of cis-jasmone-induced resistance in *Arabidopsis thaliana* and its role in indirect defence. *Planta* 232: 1163-1180. - doi: [10.1007/s00425-010-1244-4](https://doi.org/10.1007/s00425-010-1244-4)
- McFrederick QS, Kathilankal JC, Fuentes JD (2008). Air pollution modifies floral scent trails. *Atmospheric Environment* 42: 2336-2348. - doi: [10.1016/j.atmosenv.2007.12.033](https://doi.org/10.1016/j.atmosenv.2007.12.033)
- Michelozzi M, Tognetti R, Maggino F, Radicati M (2008). Seasonal variations in monoterpene profiles and ecophysiological traits in Mediterranean pine species of group "halepensis". *iForest* 1: 65-74. - doi: [10.3832/ifor0206-0010065](https://doi.org/10.3832/ifor0206-0010065)
- Pasqua G, Monacelli B, Manfredini C, Loreto F, Perez G (2002). The role of isoprenoid accumulation and oxidation in sealing wounded needles of Mediterranean pines. *Plant Science* 163: 355-359. - doi: [10.1016/S0168-9452\(02\)00139-5](https://doi.org/10.1016/S0168-9452(02)00139-5)
- Sharkey TD, Yeh S, Wiberley AE, Falbel TG, Gong D, Fernandez DE (2005). Evolution of the isoprene biosynthetic pathway in Kudzu. *Plant Physiology* 137: 700-712. - doi: [10.1104/pp.104.054445](https://doi.org/10.1104/pp.104.054445)
- Singsaas EL, Lerdau M, Winter K, Sharkey TD (1997). Isoprene increases thermotolerance of isoprene-emitting species. *Plant Physiology* 115: 1413-1420. [online] URL: <http://www.ncbi.nlm.nih.gov/pubmed/12223874>
- Staudt M, Mir C, Joffre R, Rambal S, Bonin A, Landais D, Lumaret R (2004). Isoprenoid emissions of *Quercus* spp. (*Q. suber* and *Q. ilex*) in mixed stands contrasting in interspecific genetic introgression. *New Phytologist* 163: 573-584. - doi: [10.1111/j.1469-8137.2004.01140.x](https://doi.org/10.1111/j.1469-8137.2004.01140.x)
- Steeghs M, Bais HP, de Gouw J, Goldan P, Kuster W, Northway M, Fall R, Vivanco JM (2004). Proton-transfer-reaction mass spectrometry (PTR-MS) as a new tool for real time analysis of root-secreted volatile organic compounds (VOCs) in *Arabidopsis thaliana*. *Plant Physiology* 135: 47-58. - doi: [10.1104/pp.104.038703](https://doi.org/10.1104/pp.104.038703)
- Strauss SY, Rudgers JA, Lau JA, Irwin RE (2002). Direct and ecological costs of resistance to herbivory. *Trends in Ecology and Evolution* 17: 278-285. - doi: [10.1016/S0169-5347\(02\)02483-7](https://doi.org/10.1016/S0169-5347(02)02483-7)
- Tholl D, Sohrabi R, Huh JH, Lee S (2011). The biochemistry of homoterpenes - Common constituents of floral and herbivore-induced plant volatile bouquets. *Phytochemistry* 72: 1635-1646. - doi: [10.1016/j.phytochem.2011.01.019](https://doi.org/10.1016/j.phytochem.2011.01.019)
- Velikova V, Fares S, Loreto F (2008). Isoprene and nitric oxide reduce damages in leaves exposed to oxidative stress. *Plant Cell and Environment* 31: 1882-1894. - doi: [10.1111/j.1365-3040.2008.01893.x](https://doi.org/10.1111/j.1365-3040.2008.01893.x)
- Velikova V, Varkonyi Z, Szabo M, Maslenkova L, Nogues I, Kovacs L, Peeva V, Busheva, M, Garab G, Sharkey TD, Loreto F (2011). Increased thermostability of thylakoid membranes in isoprene-emitting leaves probed with three biophysical techniques. *Plant Physiology* 157: 905-916. - doi: [10.1104/pp.111.182519](https://doi.org/10.1104/pp.111.182519)
- Velikova V, Sharkey TD, Loreto F (2012). Stabilization of thylakoid membranes in isoprene-emitting plants reduces formation of reactive oxygen species. *Plant Signaling and Behaviour* 7: 139-141. - doi: [10.4161/psb.7.1.18521](https://doi.org/10.4161/psb.7.1.18521)
- Vickers CE, Gershenzon J, Lerdau MT, Loreto F (2009). A unified mechanism of action for volatile isoprenoids in plant abiotic stress. *Nature Chemical Biology* 5: 283-291. - doi: [10.1038/nchembio.158](https://doi.org/10.1038/nchembio.158)
- Wilson ID, Neill SJ, Hancock JT (2008). Nitric oxide synthesis and signalling in plants. *Plant Cell and Environment* 31: 622-631. - doi: [10.1111/j.1365-3040.2007.01761.x](https://doi.org/10.1111/j.1365-3040.2007.01761.x)